

Histochemical Mapping of Glycoconjugates in the Eyeball of the One Humped Camel (*Camelus Dromedarius*)

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ABSTRACT

In the present study, the distribution of various sugar residues in the eyeball tissues of sexually mature camels was examined employing fluorescein isothiocyanate- (FITC) conjugated lectins. Our results revealed the presence of mannose (labeled by lectins ConA), galactose (labeled by PNA, GSAI, ECA), GalNAc (labeled by SBA, VVA), and GlcNAc (labeled by WGA) residues in the camel ocular tissues. The epithelium and stroma of the ocular tissues were labeled with mannose (ConA) and GlcNAc (WGA) binding lectins. Binding sites for WGA and PNA to the rod and cone cells of the retina were evident. The lectins Con A, WGA and GSAI bound strongly to the endothelium of blood vessels and to smooth muscle cells of the iris. In conclusion, the findings of the present study clearly indicate that the camel eyeball contains a wide range of glycoconjugates (bearing mannosyl, galactosyl and glucosyl residues), and they lack fucosyl residues.

Keywords: eyeball, camels, glycoconjugates, galactosyl, glucosyl.

INTRODUCTION

Although the camel plays an important role as a domesticated mammal in the arid regions of Africa, Asia and Australia, many aspects of its reproduction are still unknown (Zayed et al., 1995).

In recent years, lectin histochemistry has developed a useful tool to study various aspects of cell differentiation and cell-to-cell interaction (To'pfer-Petersen, 1999; Gabius, 2001); As well as the cell surface saccharides believed to be involved in a variety of cell functions, including development, growth regulation and cellular locomotion (Hakomori, 1981, Gabius et al., 1988, 2001).

It is significant that most plant and animal lectins have been classified into a rather limited number of carbohydrate-binding groups (Goldstein and Poretz, 1986). These include the mannose/glucose-binding lectins, the galactose-binding lectins, the N-acetylgalactosamine-binding lectins, the N-acetylglucosamine-binding lectins, the L-fucose-binding lectins, sialic acid-binding lectins and lectins with complex carbohydrate-binding sites. In the field of ocular tissue, lectins applied in for various purposes to study the differences between normal and migrating corneal epithelium (Gipson et al., 1983), to demonstrate the importance of glycoconjugates in the morphogenesis of the corneoscleral angle where they provide some of the required signals for the differentiation of the trabecular meshwork (Beauchamp et al., 1985). Lectins are at light microscopical level to investigate normal cornea in a variety of species (Holmes et al., 1985; Panjwani et al., 1986a; Panjwani and Baum, 1988; Tuori et al., 1994, Aly, 2003). In addition, using this technique, changes in patterns of glycosylation have been observed in wounded (Gipson et al., 1983; Gordon and Marchand, 1990) and dystrophic corneas (Panjwani et al., 1986b, 1987; Panjwani and Baum, 1989; Bishop et al., 1991), also to determine the distribution of carbohydrate residues on photoreceptor cell surfaces (Bridges, 1981, Aly, 2003). Thus, Blanks and Johnson (1984) could demonstrate selective binding-sites for lectins in the developing retina, as well as specific binding of peanut lectin to certain photoreceptor cells (Johnson and Hageman 1987). These lectin binding patterns of the ciliary body epithelium suggest a topographical and functional difference in this double cell-layered epithelium (Chan et al., 1999). Many studies on monkey, pig, cat and rabbit, eye tissues were intensely labeled with (Kawano et al., 1984).

Lectins used in eye research, mainly for human material (Prause, 1991). Lectin histochemistry is considered to be a valuable method for determining changes in the glycoconjugate content during normal and pathological conditions .

To our knowledge, no data are available concerning the camel. Therefore, the aim of present work was to determine the changes in the sugar residues within the camel eyeball by means of glycohistochemical (lectin histochemical) methods.

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MATERIALS AND METHODS

Samples

The present study was performed on the eyeball of 10 sexually mature and apparently healthy camels. Specimens were taken from the cornea, sclera, iris, ciliary body, choroid, retina and optic nerve, with a side length of 0.3 - 0.5 cm.

All samples were collected within 30 min of slaughter in a local (Cairo) abattoir. Small samples of the eyeball tissue (0.5–1 cm) were fixed in Bouin's fluid for 24 h. Thereafter, fixed samples were extensively washed in 70% ethanol (3 – 24 h) to elute fixative before tissue processing to paraffin wax by routine methods. Using a Leitz rotatory microtome (type 1521), 5-mm-thick sections were cut and mounted on both 3-aminopropyltriethoxysilane- coated and uncoated glass slides. Paraffin wax embedded sections that were kept in an incubator at 40 °C until used for glycohistochemical analysis.

Lectin histochemistry

Distribution of sugar moieties (glycoconjugates) in the adult camel eyeball tissues were investigated using 9 different fluorescein isothiocyanate- (FITC) conjugated lectins, (all purchased from Sigma Aldrich Chemicals GmbH, Deisenhofen, Germany) listed in Table 1. The lectins were chosen to represent five groups: mannose-, galactose-, N-acetylgalactosamine

(GalNAc)-, N-acetylglucosamine (GlcNAc)- and fucose-binding lectins. Lectin binding was revealed as follows: Sections were dewaxed (2 – 30 min) in xylene, rehydrated through descending grades of ethanol and washed under tap water for 10 min. They were then washed (3– 5 min) in 0.05M Tris-buffer, pH 6.8 and then incubated with 33 mg/ml FITC-conjugated lectin in Tris buffer in a humid chamber, at 4 °C, overnight.

Sections were then again washed under tap water for 5 min and subsequently rinsed (3 – 5 min) in Tris buffer (pH 6.8). Importantly, the hydrated sections were then taken directly from Tris-buffer and mounted with a 25:140 mixture of polyvinyl alcohol and ethylene glycol (Serva, Heidelberg, Germany) in Tris-buffer, pH 6.8. Mounted slides were stored at – 20 °C until examined using a fluorescent microscope.

Controls

Control sections were treated as described previously except that the FITC-conjugated lectins were either (1) substituted with Tris-buffer, or (2) pre-incubated with 0.4M of the corresponding hapten sugar inhibitor listed in Table 1 (Sigma, Deisenhofen, Germany) for 1 h before labeling.

Analysis of labeling Lectin-labeled eyeball tissues and their controls were evaluated using a Dialux 20 fluorescent microscope (Leitz GmbH, Wetzlar). Photomicrographs were captured using Kodak film elite 400.

Table 1: FITC-labeled lectins used for investigation of sugar moieties in the camel ocular tissue

Lectin group	Lectin source (Latin name)	Common name	Acronym	Sugar specificity	Binding inhibitor
I. D-Mannose (D-Glucose)-binding lectins	<i>Canavalia ensiformis</i> Agglutinin	Jack bean	Con A	α -D-Man > α -D-Glc	Man
II. D-Galactose-binding lectins	<i>Arachis hypogaea</i> Agglutinin	Peanut	PNA	β -D-Gal-(1-3)-D-GalNAc	Gal
	<i>Griffonia simplicifolia I</i> Agglutinin	Griffonia or Bandeiraea	GSA-I	Terminal α -Gal	Gal
	<i>Erythrina Cristagalli</i> Agglutinin	Coral tree	ECA	α -D-Gal-(1-4)-GlcNAc	Gal
III. N-acetyl-D-galactosamine (GalNAc)-binding lectins	<i>Glycine max</i> Agglutinin	Soybean	SBA	D-GalNAc	GalNAc
	<i>Vicia villosa</i> Agglutinin	Hairy vetch	VVA	D-GalNAc	GalNAc
IV. N-acetyl-D-glucosamine (GlcNAc)-binding lectins	<i>Triticum vulgare</i> Agglutinin	Wheat germ	WGA	GlcNAc(β 1-4GlcNAc) ₁₋₂ , NeuNAc	GlcNAc
V. L- Fucose-binding lectins	<i>Ulex europaeus -I</i> Agglutinin	Gorse seed	UEA-I	α -L-Fuc	α -L-Fuc
	<i>Lotus tetragonolobus</i> Agglutinin	Asparagus pea	LTA	α -L-Fuc	α -L-Fuc

Man, mannose; Glc, glucose; Gal, galactose; GalNAc, N-acetylgalactosamine; GlcNAc, N-acetylglucosamine; NeuNAc, N-acetylneuraminic acid (sialic acid); α -L-Fuc, α -L-Fucose.

RESULTS

Slight differences in lectin labeling to reveal different sugar moieties were seen in different ocular tissues. The results are summarized in Table 2 and described in detail below.

1- Cornea

As in other species the camel cornea is composed of five layers, the corneal epithelium, subepithelial basement membrane (Bowman's membrane), substantia propria or stroma, posterior limiting membrane (Descemet's membrane), and posterior epithelium (corneal endothelium).

Con A, WGA and ECA bound to all layer of the cornea, binding of LTA, SBA, VVA, PNA, GSA I and UEA I was observed only in epithelial layer of the cornea (Table 2).

Con A, WGA, ECA and UEA-I reacted strongly with the corneal epithelium, while the others lectin reaction were very weak (Fig.1).

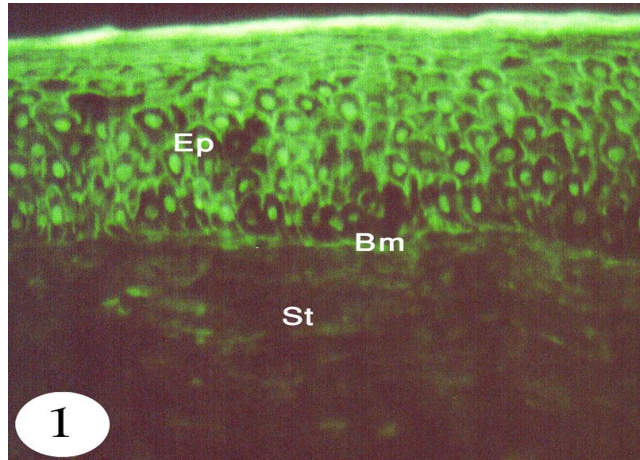


Fig. 1: Labelling of the epithelium (Ep), Bowman's membrane (Bm) and stroma (St) of the camel cornea with WGA ($\times 600$).

The binding of the Con A, WGA and PNA to the Bowman's membrane were strong (Fig.2) while the others did not bind significantly.

The corneal stroma, Descemet's membrane and endothelial cells of the cornea reacted mainly with Con A and WGA (Fig.3). Other lectins did not show any reaction or gave only a weak reaction (Fig.4).

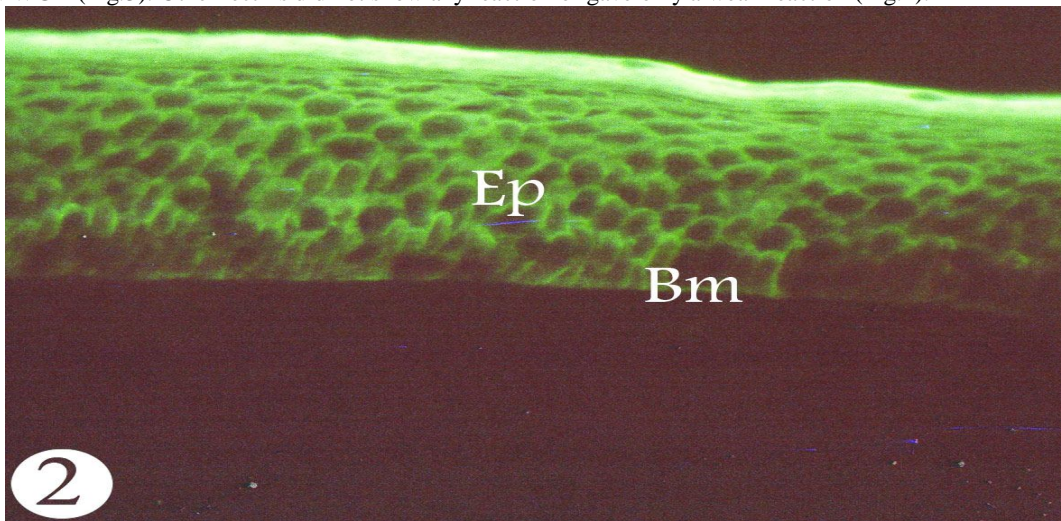


Fig. 2: Labelling of the epithelium (Ep) and Bowman's membrane (Bm) of the camel cornea with PNA ($\times 600$).

2- Sclera

The sclera consists of flat ribbons of collagenous bundles run in various directions. Between these bundles are fine elastic nets, fibroblasts and occasional melanocytes.

As in the other species, the camel sclera can be subdivided into three layers; the outermost layer, the episcleral tissue, consists of loose fibroelastic tissue. In the middle layer, the sclera proper (substantia propria), bundles of collagenous fibers are oriented mainly parallel to the surface but with some interweaving. Fine bundles of smooth muscle fibers can be demonstrated. The innermost layer, termed the lamina fusca or dark layer, is composed of much smaller bundles of collagenous fibers. Between the fibers are branching chromatophores containing melanin.

The lectins ECA and WGA bound only to the connective tissue of the sclera (Table 2).

3 - Iris

Histologically the camel iris consists of three layers: an anterior epithelial layer (endothelial layer), a middle layer of connective tissue stroma, which contains the two smooth muscles (dilator and sphincter pupillae muscles), and the posterior layer of the pigmented epithelium.

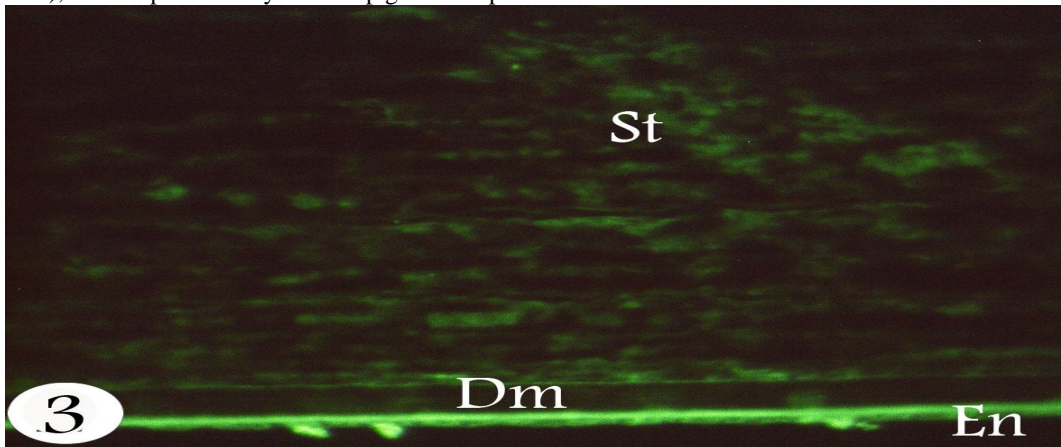


Fig. 3: Labeling of the stroma (St), Descemet's membrane (Dm) and endothelium (En) of the camel cornea with Con A ($\times 600$).

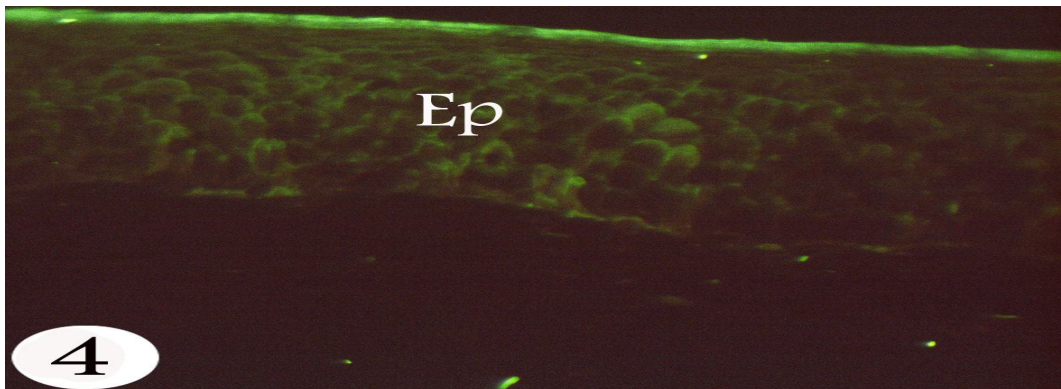


Fig. 4: Weak lablling of the epithelium (Ep) of the camel cornea with GSAI ($\times 600$).

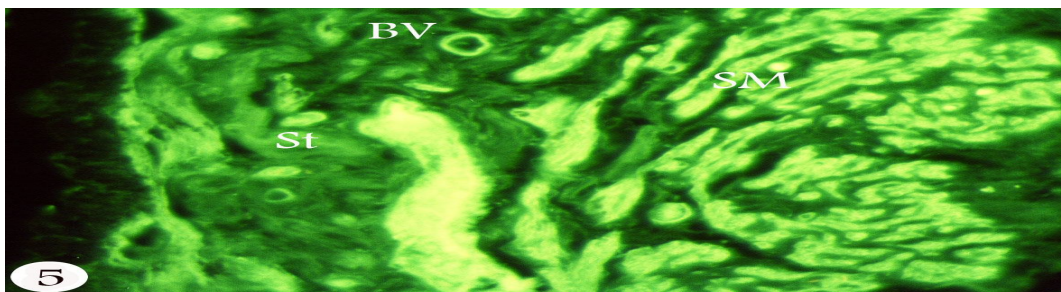


Fig. 5: Lablling the endothelium of the blood vessels (BV), iridal sphincter musscle (SM) of the stroma (St) of the camel iris with Con A ($\times 600$).

Con A, WGA, VVA and ECA reacted strongly with the stroma of the iris (Fig. 5). The binding was mostly to the connective tissue fibre. The binding of the lectin to the different layers of blood vessels varied depending on the lectin used. Con A and WGA lectin stained the two inner most layers of the tunica intima and the tunica media (Fig. 6). While the VVA and ECA bound to all layers of the blood vessels, but the staining was not particularly prominent. GSA I agglutinin bound strongly to the endothelium of all blood vessels. The reaction appeared very distinct because the adjacent stroma was negative.

Con A, WGA and VVA bound strongly to the sphincter iridal muscles (Fig. 7). Also ECA bound to it but to lesser degree. Con A and ECA bound strongly to the dilator muscle of the iris. Other lectins did not show any reaction with iridal muscles (Fig.8).

Con A and WGA reacted with the posterior pigmented epithelium of the iris. The reaction was not very strong. Other lectins did not stain the pigmented epithelium at all (Table 2).

4 - Ciliary Body

The camel ciliary body consists of the following layers; the supraciliaris layer which is the most peripheral layer of the ciliary body, the ciliary muscle, the stroma of the ciliary body that contains a large number of blood vessels, arteries and veins, the Bruch's membrane, the pigmented epithelium layer which consists of simple cuboidal or low columnar cells with rounded nuclei and the nonpigmented epithelial layer that is the internal cellular lining of the ciliary body.

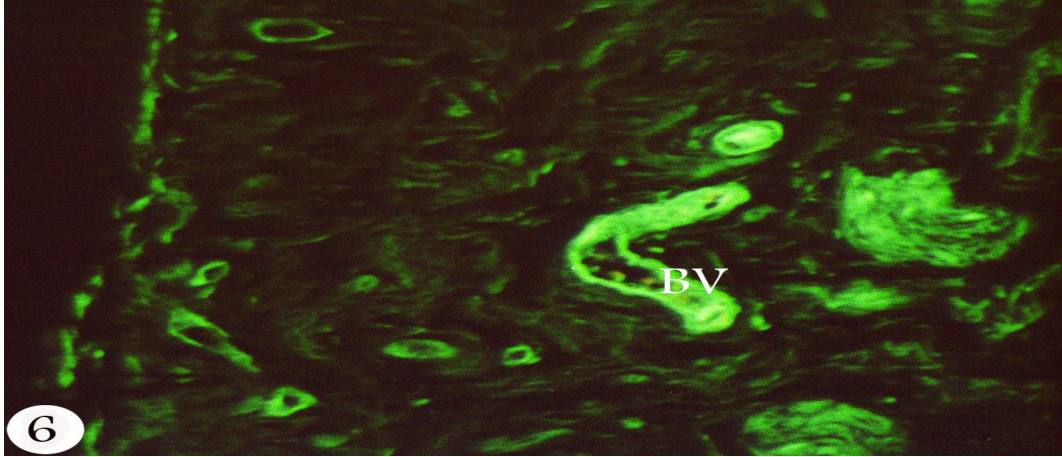


Fig. 6: Labelling the endothelium of the blood vessels (BV) of the stroma of the camel iris with WGA ($\times 600$).

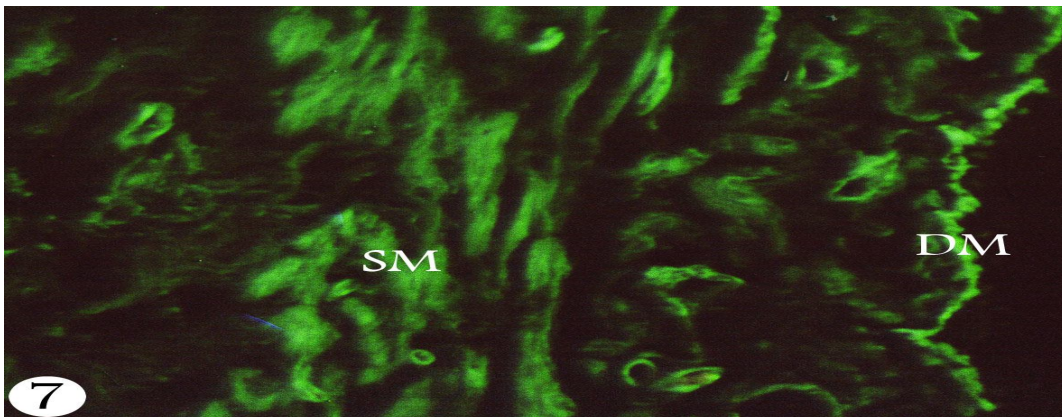


Fig. 7: Labelling the endothelium of the blood vessels (BV) and iridal dilator muscle (DM) of the stroma of the camel iris with WGA ($\times 600$).

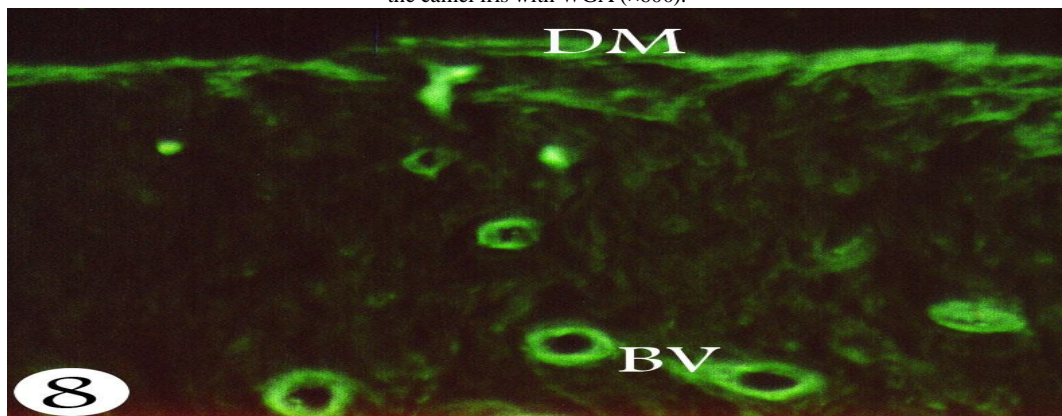


Fig. 8: Labelling the endothelium of the blood vessels (BV) and iridal dilator muscle (DM) of the stroma of The camel iris with ECA ($\times 600$).

Ciliary body of material reacted strongly with Con A and WGA (Fig. 9) while ECA and PNA are give moderate reaction. The other lectins did not give any positive reaction with samples (Table 2). Con A, WGA and PNA also labelled the smooth muscle cell of the ciliary muscles (Fig.10). The nonpigmented layer of the ciliary

epithelium bound all lectins except UEA I. Con A, WGA, and PNA showed a stronger binding of different layer of blood vessels than the other lectins (Fig.11).

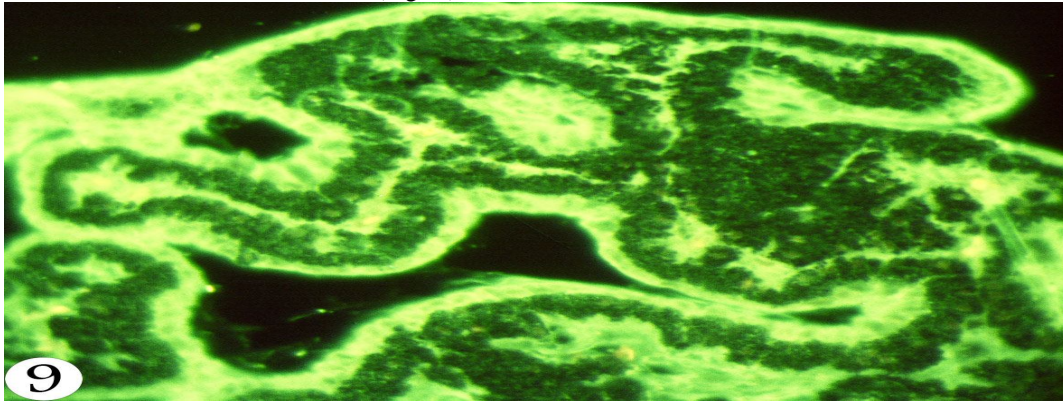


Fig. 9: Labelling the camel ciliary body with Con A (×600)

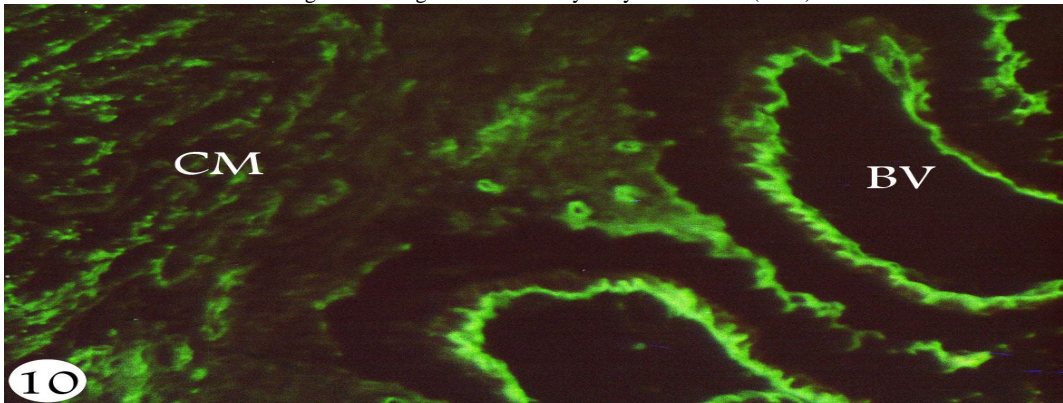


Fig.10: Labelling the endothelium of the blood vessels (BV)and ciliary muscle (CM) of the camel ciliary body with PNA (×600).

5 – Choroid

The choroidea can be subdivided into four layers as follows: the suprachoroid layer, the perichoroidal spaces, and the vessel layer which consists of intercrossing of large and medium sized arteries and veins, separated by loose connective tissue stroma rich in chromatophores.

Basal lamina and blood capillaries of the choroids reacted strongly with Con A, ECA, VVA and WGA. PNA (Fig. 12). GSA I bound strongly to all capillaries. Other lectins were not reacting with the choroid (Table 2 and Fig. 13).

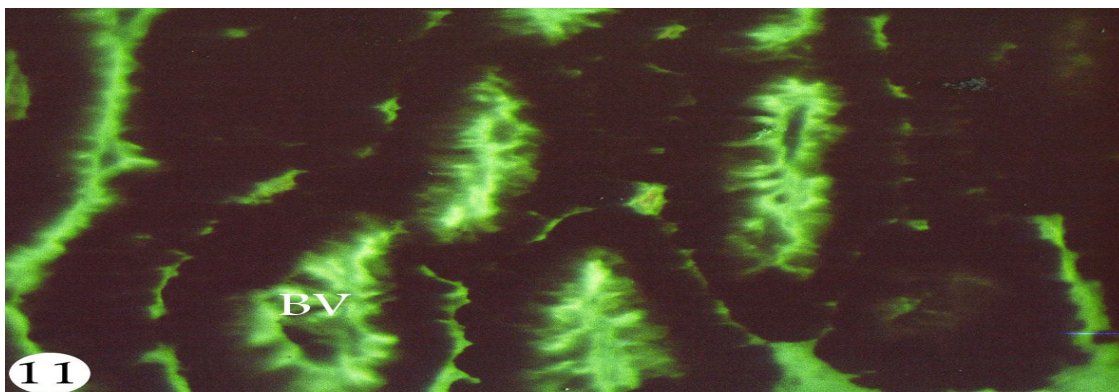


Fig.11: Labelling the endothelium of the blood vessels (BV) of the camel ciliary body with WGA (×600).

6 - Retina

Histologically, the retina consists of following layers: pigment epithelium, layers of rods and cones, external limiting membrane, outer nuclear layer, outer plexiform layer, inner nuclear layer, inner plexiform layer, ganglion cell layer, optic nerve fiber layer, and internal limiting membrane.

WGA and Con A bound strongly to all layers of the retina, mainly to the rods and cones (Fig. 14). LTA, SBA and UEA I did not give any reaction with retina.

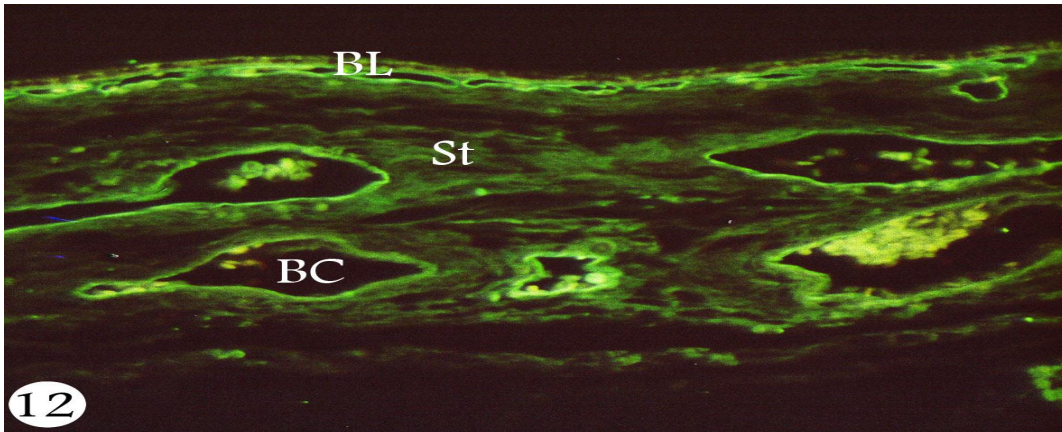


Fig.12: Labelling of the camel choroid layers; the endothelium of the blood capillaries (BC), stroma (St) and basal lamina (BL) with PNA ($\times 600$).

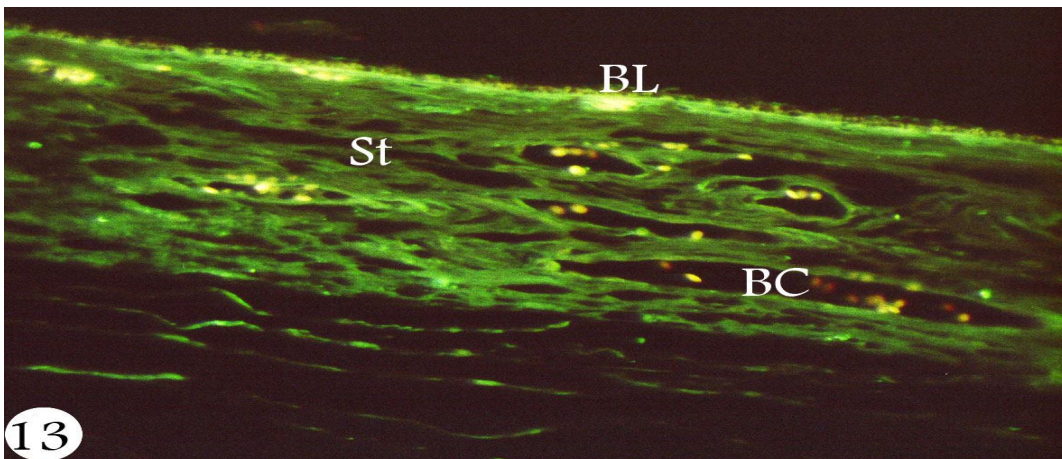


Fig.13: Labelling of the camel choroid layers; the endothelium of the blood capillaries (BC), stroma (St) and basal lamina (BL) with GSA I ($\times 600$).

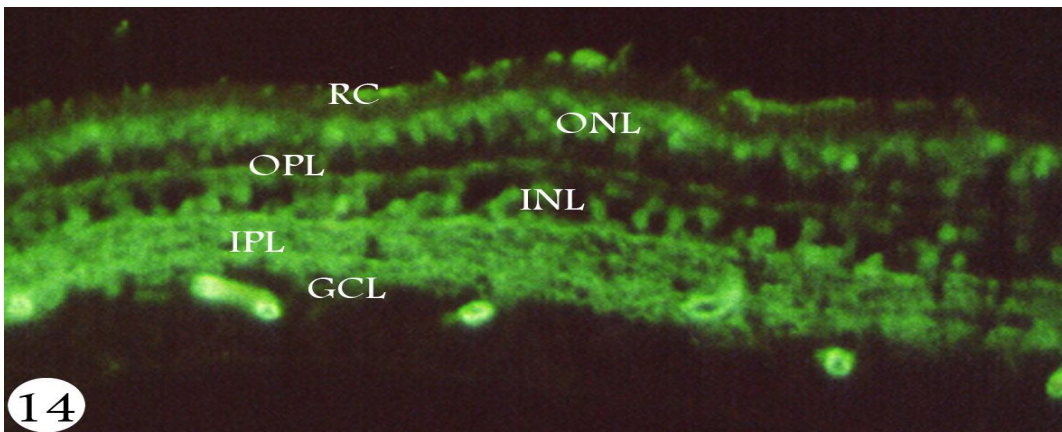


Fig.14:labelling of the layer of rods and cones (RC), outer nuclear layer (ONL), outer plexiform layer (OPL), Inner nuclear layer (INL), inner plexiform layer (IPL) and Ganglion cell layer (GCL) of the camel retina with WGA ($\times 600$).

In detail, the retina showed following staining pattern: RPE revealed strong lectin binding sites for Con A, VVA WGA, and PNA (Fig. 15), while GSA I agglutinin showed a weak reaction with RPE. Other lectins did not give any positive reaction with RPE. WGA, Con A, VVA and PNA bound strongly to the rod and cone photoreceptor cells. GSA I agglutinin reacted only strongly with the outer segment of the rods and cones.

Con A, WGA, and PNA bound strongly to the external and internal limiting membrane of the retina (Table 2), but the reaction was more prominent in external limiting membrane. Other lectins did not give any staining, neither of the external or the internal limiting membrane. Whereas WGA and Con A reacted weakly with both

outer and inner nuclear layers of the retina. Con A was the only lectin which showed moderate binding to the ganglionic cell layer in retina. All others did not give any reaction.

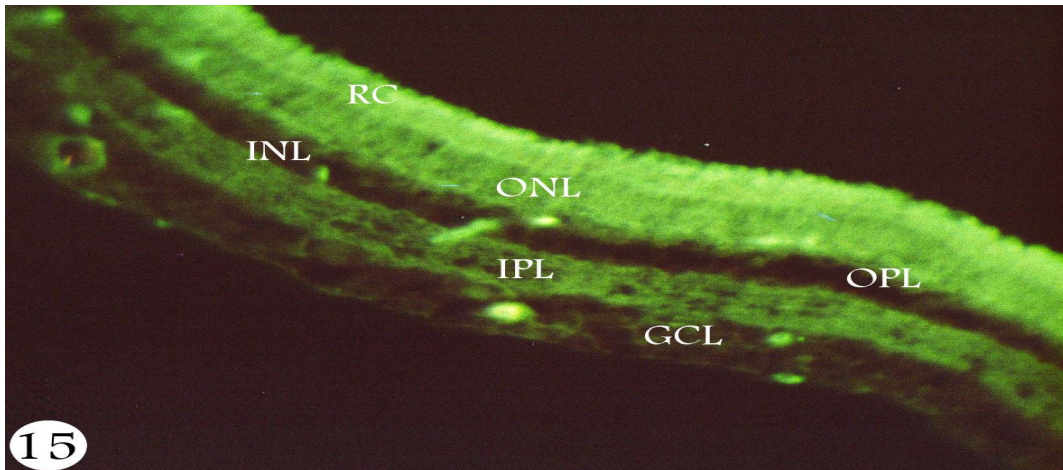


Fig. 15:labelling of the layer of rods and cones (RC), outer nuclear layer (ONL), outer plexiform layer (OPL), Inner nuclear layer (INL) and inner plexiform layer (IPL) and of the camel retina with PNA ($\times 600$). GCL = Ganglion cell layer

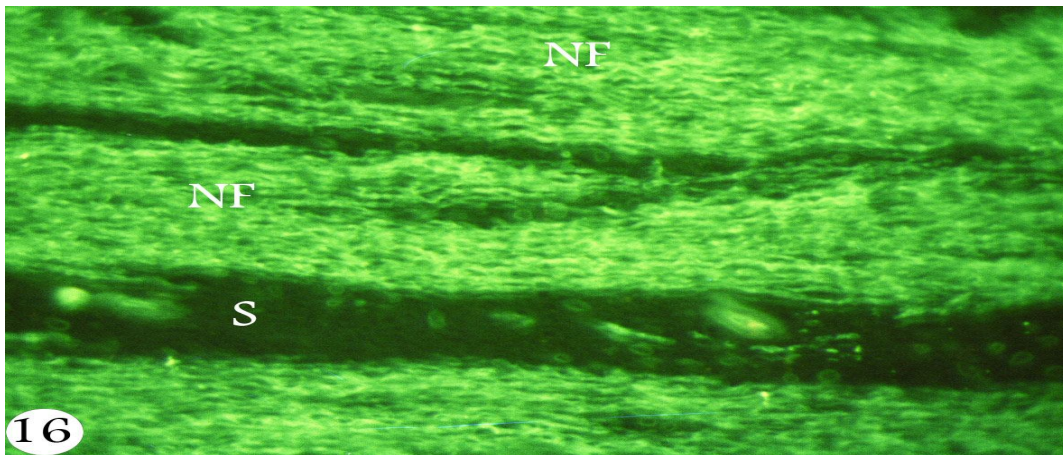


Fig.16: Labelling of the nerve fiber (NF) of the camel optic nerve with VVA ($\times 600$).
S= Connective tissue septa

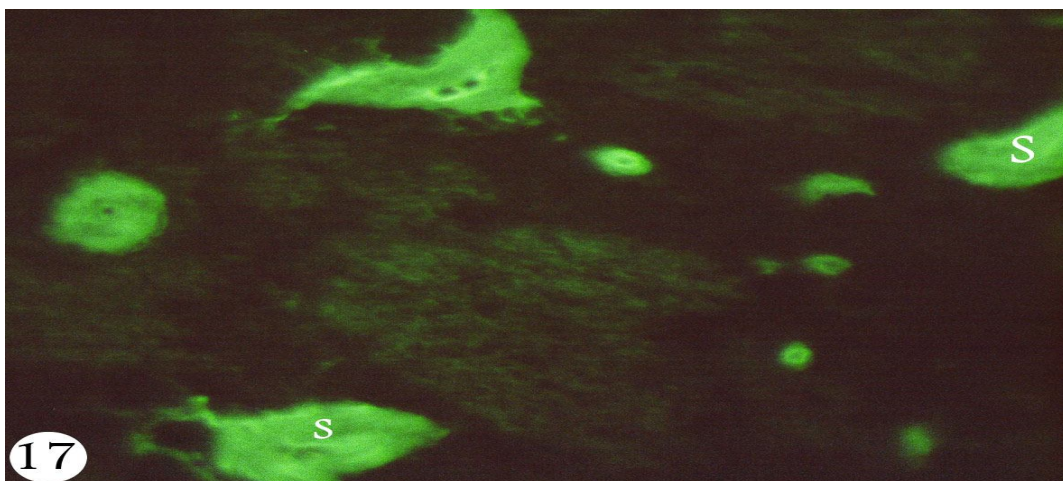


Fig.17: Labelling of the connective tissue septa (S) of the camel optic nerve with WGA ($\times 600$).

WGA, Con A, VVA and PNA reacted weakly with the optical nerve fiber layer of the retina (Fig. 15). GSA I agglutinin reacted strongly with the endothelium of all blood vessels in the retina. Con A, VVA, WGA, and PNA showed the endothelium of the retinal blood vessels weakly. All other lectins were negative (Table 2)

7 - Optic Nerve

The optic nerve is formed by ganglion cell axons, glial cells and septa of connective tissue which arise from the pia mater. The area cribrosa is formed by lamellae of collagenous fibers which, run in different directions forming a mesh like arrangement and are penetrated by the axons of the optic nerve.

In sagittal sections the bundles of the fibers of the optic nerve run parallel to each other separating by collagenous bundles septa of the area cribrosa. These bundles fade out in posterior directions.

Nerve fibers of the optic nerve bound strongly ECA, VVA, PNA and UEA I (Fig. 16).. There was no positive reaction with Con A, WGA, LTA, SBA and GSA I (Fig. 17).

Bundles of the collagen fibers of the connective tissue septa in the area cribrosa, reacted strongly with Con A, WGA and ECA (Fig. 17). They showed a weak reaction with LTA and PNA and did not stain with VVA, SBA, GSA I and UEA I. Collagenous fibers in the area cribrosa. The staining with ECA of the collagenous fibers from the area cribrosa was not particularly prominent. Blood capillaries of the optic nerve, reacted strongly with Con A, ECA, SBA, WGA and GSA I (Fig. 17) while bounded weakly VVA, and no reaction was seen with LTA, PNA and UEA I.

Table 2: lectin binding sites in the camel eyeball fixed with Bouin's solution.

Sites	Con A	LTA	ECA	SBA	VVA	WGA	PNA	GSA I	UEA I
Cornea									
Ep	+++	+	+	+/-	+/-	+++	+++	+	++
Bm	++	-	-	-	-	++	++	-	-
St	++	-	+/-	-	-	++	+/-	-	-
Dm	++	+/-	+	-	+/-	++	+/-	-	-
En	++	+	+/-	+/-	+/-	++	+/-	-	-
Sclera	-	-	+	-	+/-	+/-	-	-	-
Iris									
PE	+++	-	+/-	-	-	+++	-	-	-
St	++	-	++	-	++	++	+	-	+
BV	+/++	+/-	+/++	+/-	++	+/++	++	+++	-
SM	++	-	+	-	+/++	++	-	-	-
Ciliary body									
CM	+	-	-	-	-	+	+	-	-
NPE	++	+	+	+	+	++	++	+	-
Choroid									
BL	++	-	++	-	++	++	-	-	-
BV	++	-	++	-	++	++	++	++	-
Retina									
RPE	+/+++	-	-	-	+/+++	+/+++	+/+++	+/-	-
R&C	+/+++	-	-	-	+/+++	+/+++	+/+++	++	-
ELM	++	-	-	-	-	++	++	-	-
ONL	+	-	-	-	-	+	-	-	-
OPL	+	-	-	-	-	+	+	-	-
INL	+	-	-	-	-	+	-	-	-
IPL	+	-	-	-	-	+	+	-	-
GCL	+	-	-	-	-	-	-	-	-
NFL	+	-	-	-	+	+	+	-	-
ILM	++	-	-	-	-	++	++	-	-
BV	+	-	-	-	+	+	+	+++	-
Optic nerve									
NF	-	+	+	-	+	-	+	-	+
S	++	+	++	-	-	++	+	-	-
BV	++	-	++	++	+	++	-	++	-

- = negative reaction + = week reaction ++ = moderate reaction +++ = strong reaction

Ep = Epithelium, Bm = Bawman's membrane, St = Stroma, Dm = Descemet's membrane, En = Endothelium, PE = Pigmented epithelium, NPE = Non pigmented epithelium, BV = Blood vessels, SM = Sphincter muscle, CM = Ciliary muscle, BL = Basal lamina, RPE = Retinal pigmented epithelium, R&C = Rods and cones, ELM = External limiting membrane, ONL = Outer nuclear layer, OPL = Outer plexiform layer, INL = Inner nuclear layer, IPL = Inner plexiform layer, GCL= Ganglion cell layer, NFL = Nerve fiber layer, ILM = Internal limiting membrtane, S = Septa, NF = Nerve fiber.

DISCUSSION

Glycoproteins occur mainly intracellular and may also be found in cell membranes with a variety of important biological functions. Lectins are sugar binding protein that can be useful for the localization of glycoproteins in cells. This could contribute to a better interpretation of the physiological and pathological processes in the corneal tissue (Bonvicini et al., 1983).

Post-translational glycosylation of proteins and lipids play an important role for the cellular functions. Lectin histochemistry enables the morphological evaluation of the distribution of the saccharide residues within the tissue sections (Spicer and Schulte, 1992). Thus the present data, demonstrating the lectin binding sites in the camel eyeball, propose a basis for further analyses of the role of saccharide residues in the eye under different experimental and pathological conditions.

The pattern of lectin binding in the corneal epithelium suggests the presence of glycoconjugates containing terminal α -mannose, N-acetylglucosamine and sialic acid residues and sparse terminal α -galactose and β -N-acetylgalactosamine residues (Panjwani et al., 1986a; Rittig et al., 1990; Bishope et al., 1991 and Lawrenson et al., 1998). This generally agree with our results.

Lectin binding to the cornea in different species, e.g. calf has been studied previously by Panjwani and Baum, 1989. The results of this study are generally in accordance with their findings but there are also some differences: Panjwani and Baum, 1989 mentioned that Con A and PNA binds to the corneal epithelium, whereas in our results show that Con A binds throughout all epithelial layers of the cornea. Also PNA was bounded mainly to the apical epithelial cells (Panjwani and Baum, 1989) rather than to basal cells as demonstrated in our study. Our results are different to the findings of Tuori et al., 1994 who reported no binding also of the Con A and PNA to the corneal epithelium. Tuori et al., 1994. demonstrated that WGA, UEA-I and GSA-I bound to the corneal epithelium and that apical cells of the epithelium of the cornea displayed more α -GalNAc, GlcNAc, sialic acid and α -L-Fuc residues than the basal cells. This agrees with our results. The different staining pattern between apical and basal cells of the corneal epithelium has been described previously by several authors in different species (Gipson et al., 1983; Bonvicini et al., 1983; Panjwani et al., 1986a; Rittig et al., 1990), and it is related to the differentiation of epithelial cell, as they move to the apical layers of the epithelium (Nemanic et al., 1983).

Furthermore, Con A and WGA bound to the corneal stroma in our experiment, but no binding of this lectins was seen in the study of Panjwani and Baum, 1989 who reported that PNA is the only lectin that binds to the corneal stroma. This result go to with findings of Tuori et al., 1994 and our results.

Our findings on corneal tissue do not parallel those of (Spiro and Bhoyroo, 1984), but show excellent correlation with results of Panjwani and Baum, 1988. The observed species differences in the expression of the corneal stromal GSA-I binding sites probably do not reflect differences in blood group antigens among different species, because, at least in humans, such antigens are not found in stromal matrix or on stromal cells (Herold, 1972).

Con A and WGA bound to the Descemet's membrane of the camel cornea. This result is generally in accordance with the studies of Panjwani and Baum, 1989 and Tuori et al., 1994, but with a little difference. Con A reacted with the anterior part of the Descemet's membrane, while WGA reacted mainly with the posterior border of the Descemet's membrane. Heterogeneity in the distribution of the glycoproteins within the Descemet's membrane has been demonstrated previously (Gordon, 1990; Ljubimov et al., 1995 and Lawrenson et al., 1998). The thin basement membranes showed a marked presence of N-acetylgalactosamine residues, whereas a low concentration of these sugar residues was found in thick basement membranes (Salamat et al., 1993).

Our observations showed binding of Con A and WGA to the corneal endothelium. This finding is not in accordance with the results of Panjwani and Baum, 1989 and Tuori et al., 1994 who reported that the corneal endothelium tend to bind only GSA-I-B4.

Some of the results from previous lectin binding studies in the human cornea are somewhat different in various investigations (Bonvicini et al., 1983; Panjwani et al., 1986a; Brandon et al., 1988 and Bishop et al., 1991).

Brandon et al., 1988 suggested that some of the variation in the histochemical lectin binding studies is due to different staining procedures or post-mortem changes of the tissues. This could also explain the difference observed between our investigation and the studies of Panjwani and Baum, 1989 and Tuori et al., 1994.

PNA is a lectin which preferentially detects α -galactose. It is also used as a biological marker to detect the T antigen. Due to the neoexpression of T antigen in malignant cells, PNA and other lectins of the same specificity have been used as tools in the diagnosis of cancer. In our work PNA reacted weakly with the camel cornea.

Contrary Con A, WGA and PNA showed a distinct reactivity with Bowman's membrane, showing α -mannose, N-acetylglucosamine and α -galactose residues in this structure.

The lectin binding to the sclera did not differ significantly to that of the stroma of the cornea.

The binding of lectins to the blood vessels of the anterior uvea demonstrates the presence of gloconjugates containing terminal N-acetylglucosamine and α -galactose in the vascular endothelium. The weak staining with these lectins revealed also the presence of some α -mannosyl, N-acetylglucosamine and sialic acid residues in the endothelium. The vascular endothelium has been previously studied using some of these lectins in bovine tissues (Alroy et al., 1987, Tuori et al., 1994). Our demonstration of the presence of α - and β - galactose and sialic acid and the absence of fucose are in agreement with the results of Alroy et al., 1987, and Tuori et al., 1994. We also found mannose residues in the endothelium similar to finding of Tuori et al., 1994 and in contrast with the results of Alroy et al., 1987. Previous studies have suggested that GSA-I-B4 is an endothelial marker in mouse tissues (Laitinen, 1987) in the same way as UEA-I is for human tissues (Holthöfer et al., 1982). The present results show that GSA-I is an endothelium marker in camel tissues and these findings are in accordance with observations of Tuori et al., 1994.

The stroma of the iris is abundant in collagen fibers. They were stained by Con A, ECA, VVA, WGA and PNA. The presence of sialic acid and β -galactose has been noticed previously by Pena et al., 1981. Similar results were also obtained by Tuori et al., 1994.

The posterior pigmented epithelial cell membrane of the iris and the nonpigmented epithelium cell of the ciliary body have α -mannose and N-acetylglucosamine residues. This is in accordance with the studies of Tuori et al., 1994.

α -mannosyl and N-acetylglucosamine residues are abundant in the camel iridal and ciliary muscle, whereas the Gal-(β 1,3)-N-GalNAc residues are also present in the ciliary muscle. Identical results were obtained by Tuori et al., 1994. Lectin histochemistry has been applied to human and rat skeletal muscle previously (Pena et al., 1981) and it has been shown that Con A reacted with muscle cells whereas UEA-I was negative. However, the other lectins used in the present study (PNA and WGA) stained iridal and ciliary smooth muscle cells differently.

The results of our study show clearly that specific structures in the camel retina can be stained with different lectins (Table). The binding of the retinal structures was dependent on the sugar-binding specificities of the different lectins, demonstrating the presence of different glycoconjugates in specialized parts of the retina.

The binding of Con A and WGA to camel retinal structures is in agreement with previous studies in the frog (Bridges, 1981), in the monkey (Uehara et al., 1983a) and in human (Söderström, 1988). However, there are some differences in the staining pattern of PNA and UEA-I that may result from species differences or from variation in tissue preparation, which are known to affect lectin histochemistry (Brasitus et al., 1982 and Söderström et al., 1984).

In our study, normal camel retinal pigmented epithelium showed lectin binding sites for Con A, WGA, VVA and PNA. These results are in accordance with the findings of (Bopp et al., 1992) in human retinal pigmented epithelium.

The biochemical role of lectin binding sites in RPE still remains unclear, especially the significance of the presence or absence of certain cellular sugar residues for structure and function of RPE-cells (Bopp et al., 1992). Con A also binds with high affinity to rhodopsin, the photoreceptor molecules of the rods (Fukuda et al., 1979; Liang et al., 1979 and Bridges and Fong, 1980). Rhodopsin contains a special oligosaccharide-chain GlcNAc β 1-2Man α 1-3(Man α 1-6) Man β 1-4GlcNAc β 1-4GlcNAc-Asn (Fukuda et al., 1979 and Liang et al., 1979). Its high content of both terminal N-acetylglucose and α -mannose residues explain the binding of Con A to the rods in camel retina. Thus, the distribution of Con A-binding sites in camel rods might reflect the distribution of rhodopsin within these cells. However, there are also other glycoconjugates in the rods, such as the outer segment protein, with a molecular weight of 291,000 (Dreyer et al., 1972; Bownds et al., 1974) and the glycoprotein of the interphotoreceptor matrix (Adler and Klucznik, 1982) that bind Con A and WGA.

Rod and cone discs are formed by infolding of the plasma membrane. In the rods these infoldings are sealed off to form stacks of flattened bimembranous discs, so that the oligosaccharide layer that normally resides on the extracellular surface (Hirano et al., 1972) is sequestered into the disc interior.

Con A stained material in the camel rods seemed to be intracellularly located and its concentration was higher in the outer segments than in the inner segments.

There are membranous disks inside the rod outer segment that contain rhodopsin (Jan and Revel, 1974 and Basinger et al., 1976). It has been suggested that the rod outer segments bind fucosyl and galactosyl residues prior to disk shedding and phagocytosis by the cells of the pigment layer (McLaughlin and Wood, 1980; O'Brien, 1976). In this way the cells of the pigment layer can recognize the differences between shed and intact disks. Previous studies have given conflicting results concerning this theory. In the monkey retina, RCA I, which identifies terminal galactosyl residues, binds to the outer segments of the rods, indicating the presence of galactosyl residues. However, in the monkey retina, UEA I and LTA which identify fucosyl residues bind only to the inner segments of the rods and cones (Bunt and Klock, 1980 and Uehara et al., 1983a). In our study we could not find any binding of ECA, UEA I or LTA in the rods. These findings are in accordance with those of Bridges, 1981 and could be due to species differences, indicating a different processing of rhodopsin in different species. However, in the rods of the frog, RCA I staining is seen on the surface of the outer segments (Bridges and Fong, 1979). Those studies were done with cell suspensions in which the cell surface is easier to study. In the present investigation tissue sections were used. The stained material is primarily intracellular. If the cell surface is only slightly stained, it may be difficult to observe it in tissue sections. Sheets and occasional chunks of this matter were sometimes found on the detached retina and probably represent the interphotoreceptor matrix, which is known to contain fucose residues (Feeney, 1973).

Since Con A and WGA also bind to the outer segment of the cones, their visual pigments may contain sugar sequences that resemble those found in rhodopsin, but show different lectin affinities. For instance, rhodopsin and iodopsin both bind to Con A columns but they are eluted at different concentrations of α -methyl-mannopyranoside (Fager and Fager, 1978).

WGA is bound to terminal GlcNAc or sialic acid residues (Goldstein and Hayes, 1978). WGA does not bind to rhodopsin (Yamamoto et al., 1983), so the binding of WGA to the retina must be explained by the presence of other glycoconjugates. These are present in the cytoplasm of the rods and cones but are not seen in the internal segment of the cones. Quite interesting is the band-like staining by WGA within the outer plexiform layer, for which no clear morphological equivalents can be seen with conventional staining methods.

Lectin PNA is known to have high affinity for α -D-galactose and N-acetylgalactosamine (Lotan et al., 1975). It is, therefore, possible to assume that these carbohydrate residues are present at the terminal oligosaccharides of membrane glycoconjugates in rods and cones. Concerning the binding sites of PNA, it is noticeable that the labelling was uniform throughout the outer and inner segments of rods and cones.

In the monkey retina, PNA also selectively stained the cones, but the binding was concentrated to the outer segment (Uehara et al., 1983b). There was also found in human, (Söderström, 1988) in pig and cat and in rabbit retina (Kawano et al., 1984). These observations are different from our results. We showed that PNA bound strongly with rods and cones layer. Our finding agrees with results which reported by Kawano et al., (1984) that mammals including rat and bovine and non-mammals including birds and goldfish strongly bound PNA to rods and cones.

In addition to the layer of the rods and cones, Con A stained many other structures of the retina. The external and internal limiting membrane was only slightly stained whereas in the outer nuclear layer, where the nuclei of the rods and cones are located. The cell surfaces of the neurons in the inner nuclear layer were marked. Both inner and outer plexiform layers were diffusely stained with Con A. This finding agrees with results in the human retina (Söderström, 1988).

The internal and external limiting membranes and the wall of the retinal vessels are labelled with PNA. The vessel wall contains collagen fibrils (Hogan et al., 1971) which are abundant in α -galactose (Muir and Lee, 1969).

In summary, our study shows that the normal camel eye contains a distinct distribution pattern for several lectins. These results on the normal camel eye may form the basis for future studies concerning changes in lectin staining occurring in different diseases of the eye.

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